

# Taq II PCR Master Mix (2X)

Cat. No.: BIO-5181-20 BIO-5181-200 BIO-5181-1000

**Concentration**:  $60U/\mu I$  Taq II DNA polymerase (recombinant), 400uM dNTP mix,  $3mM MgCl_2$ 

Storage: -20°C.

Avoid frequent thawing and freezing.

# **Quality Control**

Nuclease Assay: No detectable contaminating endonuclease or exonuclease activity.

**Functional Assay**: Taq II PCR Master Mix (2X) was tested for amplification of a 3500bp of human genomic DNA.

#### 1.0 DESCRIPTION

Taq II PCR Master Mix (2X) is a premixed, ready to use solution which contains Taq II DNA polymerase, dNTPs, MgCl<sub>2</sub> and reaction buffer for amplification of DNA templates by PCR. This pre-mixed formulation saves time and reduces contamination by reducing the number of pipetting steps required for usual PCR set up. The mix is optimized for efficient and reproducible PCR.

#### Source

An *E.coli* strain that carries a cloned *pol* gene from *Thermus aquaticus*.

#### **Unit Definition**

One unit of the enzyme catalyses the incorporation of 10nmol of deoxyribonucleotides into an acid-insoluble material in 30mins at 74°C by using activated salmon sperm DNA as a template/primer.



# Applications

- DNA sequencing
- DNA labeling
- PCR for cloning
- PCR amplification of DNA fragments up to 6kb
- Routine PCR

# Taq II PCR Master Mix (2X) Composition

Contains reaction buffer, 0.06U/ul of Taq II DNA polymerase,  $3mM MgCl_2$  and 400uM of each dNTPs.

# Remarks

- Half-life of this enzyme is >40mins @ 95°C.
- Taq II DNA Polymerase accepts modified nucleotides (e.g. biotin-, digoxigenin-, fluorescent-labeled nucleotides) as substrates for the DNA synthesis.

# 2.0 Protocol

The following protocol serves as a starting point and general guideline for any Polymerase Chain Reaction (PCR). Reaction conditions (incubation times and temperatures, concentration of Taq II DNA polymerase, primers, MgCl<sub>2</sub> and template DNA) vary and need to be optimised. PCR reactions should be prepared in a DNA-free environment, dedicated pipette and aerosol resistant tips are recommended. Always keep the control and template DNA to be amplified isolated from other components.

Recommendations of Template DNA in a 50 µl reaction volume		
DNA Type	Amount of DNA	
Human genomic DNA	0.1 to 1 µg	
Plasmid DNA	0.5 to 5 ng	
Phage DNA	0.1 to 10 ng	
<i>E.coli</i> genomic DNA	10 to 100 ng	

- 1. Thaw Taq II PCR Master Mix at room temperature. Keep the tubes on ice after thawing. Vortex briefly and spin down contents quickly.
- 2. Prepare the following reaction mix in a sterile, nuclease-free PCR tube on ice.



#### Table 1. For a 50µl reaction volume:

Components	Volume (µl)	Final Concentration
Taq II PCR Master Mix (2X)	25	1X
Forward Primer (10µM)	0.5 - 5	0.1 – 1µM
Reverse Primer (10µM)	0.5 - 5	0.1 – 1µM
DNA Template	1-5	See above
Nuclease-Free Water	Top up to 50	N.A

3. Cap tubes and spin down contents briefly.

 Place reactions in thermal cycler and incubate at 95°C to completely denature template DNA. Perform 25 - 35 cycles of PCR amplification by repeating steps 2 to 4 in the PCR reaction. Low amounts of starting template may require 40 cycles.

Step	Temperature	Time
Initial Denaturation	95°C	1 – 5 min
Denaturation	95°C	0.5 – 1 min
Annealing	50- 68°C	0.5 – 1 min
Extension	72 - 75°C	*0.5 – 1 min/kb
Final Extension	72 - 75°C	5 – 15 min
Soak	4°C	$\infty$

Table 2. PCR Amplification as follows:

\* For amplification of >3kb, it is recommended to use 1min/kb as extension rate.

# 3.0 General guidelines for PCR amplifications

# Initial denaturation

To ensure efficient utilisation of the template during the first amplification cycle, it is essential that the template is denatured completely. If GC content of the template is  $\leq$  50%, an initial 1-5 mins denaturation at 95°C would be sufficient.



# **Denaturation**

DNA denaturation time of 0.5min/cycle at 95°C is usually sufficient. For GC-rich templates, denaturation could be prolonged to 3-4mins.

# Annealing

Annealing temperature should be  $5^{\circ}$ C lower than melting temperature (T<sub>m</sub> -  $5^{\circ}$ C) of primers. 0.5min/cycle is usually sufficient. If non-specific PCR products are observed, the temperature should be optimised stepwise in 1-2°C increments.

# **Extension**

Optimal temperature for extension of Taq II DNA Polymerase is between 70-75°C. Recommended extension step is 30sec/kb at 72°C for PCR products.

#### Number of cycles

If less than 10 copies of template are present, about 40 cycles are required. For a higher amount, 25-35 cycles are sufficient.

# Final extension

After the last cycle, incubate PCR mixture at 72°C for an additional 5-15mins to allow any possible incomplete amplification to take place.