

Ver. 1.0

# PrimeWay Stool DNA Extraction Kit (KIT-9070)







# **PrimeWay Stool DNA Extraction Kit**

### Product No: KIT-9070

PrimeWay Stool DNA Extraction Kit is a robust and reliable kit designed to isolate genomic DNA from both animal and human stool sample, as well as gut content. This kit is meticulously optimised to handle broad spectrum of stool types such as normal, dry, hard & high fibre stool. Ceramic beads coupled with homogenisation duration is perfected in this kit to maximise the lysis of the microorganism. The extracted DNA is free from PCR inhibitor, making it suitable for downstream application such as PCR, amplicon sequencing, qPCR, Southern blot, etc.

For Research Use Only. Not for use in Diagnostic Procedures.

| No | Product                   | KIT-9070-10<br>10 preps | KIT-9070-50<br>50 preps |
|----|---------------------------|-------------------------|-------------------------|
| 1  | STL1 Buffer               | 9 mL                    | 50 mL                   |
| 2  | STL2 Buffer               | 1.8 mL                  | 15 mL                   |
| 3  | STL3 Buffer               | 14 mL                   | 2 x 45 mL               |
| 4  | Wash Buffer ST            | 3 mL                    | 25 mL                   |
| 5  | Elution Buffer            | 1.5 mL                  | 6 mL                    |
| 6  | Inhibitor Removal Column  | 10 pcs                  | 50 pcs                  |
| 7  | PrimeWay Stool Column     | 10 pcs                  | 50 pcs                  |
| 8  | Stool Bead Tube           | 10 pcs                  | 50 pcs                  |
| 9  | 2 mL Microcentrifuge Tube | 10 pcs                  | 50 pcs                  |
| 10 | Collection Tube           | 10 pcs                  | 50 pcs                  |

### **Kit Contents**



# Storage

This kit can be stored at room temperature (21 - 25 °C).

# **Product Specification**

|                  | KIT-9070                |
|------------------|-------------------------|
| Binding capacity | Up to 50 µg             |
| Sample Size      | Refer "Sample Material" |
| Elution          | 30 – 100 μL             |
| Duration         | ≤ 60 minutes            |

# Sample Material

✓ Sample types:

|                         | Input Amount | Example              |
|-------------------------|--------------|----------------------|
| Human stool (soft)      | 180 – 220 mg | Human                |
| Animal stool (soft) or  | 60 – 100 mg  | Dog, cat, mouse gut  |
| gut content             |              | content              |
| Dry, hard or high fibre | 60 – 100 mg  | Horse, rabbit, mouse |
| animal stool            |              |                      |

- ✓ Upon stool collection, stool samples should be kept at 2 − 8 °C if the extraction process is intended to be carried out on the same day. If DNA extraction is not performed on the same day, store the stool in −80 °C until further processing.
- ✓ Frozen stool is required to be thawed and kept on ice before DNA extraction.

## Materials Supplied by User

- ✓ Absolute ethanol (≥ 99.5 %)
- ✓ Centrifuge, at speed of 4,000 16,000 x g
- ✓ 1.5 mL microcentrifuge tubes
- ✓ 2 mL microcentrifuge tube
- ✓ 50 centrifuge tube (for hard, dry or high fibre animal stool only)
- ✓ Pipettes & pipette tips



- ✓ Vortex mixer
- ✓ Cell Disruptor (Digital Disruptor Genie, or similar instrument)
- ✓ Water bath
- ✓ Blender (for hard, dry or high fibre animal stool only)

#### **Precautions for Use**

- ✓ Stool samples should be handled in Laboratory Biosafety Level 2 (BSL-2) as it may contain pathogens such as bacteria, fungus, virus and parasite.
- ✓ Always wear a lab coat, disposable gloves, and surgical mask.
- Some buffers in this kit contain irritants. Handle with care and avoid contact with skin. In case of contact, wash skin with a copious amount of water; seek medical attention.

#### **Before Start**

- ✓ It is highly recommended to read through the manual prior to starting, especially for a first-time user.
- ✓ Ensure that no precipitation is observed in STL1 Buffer. If any precipitate is present, dissolve it by incubating the buffer at 37 °C water bath, followed by gently shaking.
- ✓ Add absolute ethanol ( $\geq$  99.5 %) to Wash Buffer ST as below:

| P/No        | Ethanol to be added |
|-------------|---------------------|
| KIT-9070-10 | 12 mL               |
| KIT-9070-50 | 100 mL              |



## Protocol

### **Reagent Supplied by User**

- ✓ [Hard, dry or high fibre animal stool only] 1X Tris-EDTA (TE) Buffer, pH 8.0
- ✓ [Optional] 100 mg/ mL RNase A

| _           | I. Preheat the Elution Buffer at 60 °C.  |  |
|-------------|--|--|
| Preparation |  |  |
|             | 1A) <u>Human stool (soft)</u>  |  |
|             | i) Transfer <b>180 – 220 mg stool</b> into the <b>Stool Bead Tube</b> . Proceed  |  |
|             | to Step 2.   |  |
|             | 1B) Animal stool (soft) or gut content   |  |
|             | Transfer 60 – 100 mg stool or gut content into the Stool Bead Tube.  |  |
|             | Proceed to Step 2.   |  |
| a           | 1C) Hard, dry or high fibre animal stool   |  |
| Sample      | <ul> <li>Add 1 volume of stool with 5 volumes of 1X TE, pH 8 into the<br/>blender.</li> </ul>  |  |
| Sar         | <ul> <li>Blend the stool with pulse mode until homogenise. This is to<br/>break the dry stool and/or fibre break into pieces and ensure<br/>uniform distribution of microbiota.</li> </ul> |  |
|             | <li>iii) Transfer the stool mixture into a new 50 mL centrifuge tube (not<br/>provided).</li>  |  |
|             | <ul> <li>iv) Centrifuge at 4,000 x g for 5 minutes. Discard supernatant.</li> <li>v) Weight 60 – 100 mg stool into the Stool Bead Tube.</li> <li>vi) Proceed to Step 2.</li> </ul>         |  |
|             | <b>Note:</b> DO NOT transfer undigested food such as crop, fruit husks and undigested seeds into the <b>Stool Bead Tube</b> .  |  |



|                   | 2.       | Add <b>800 µL STL1 Buffer</b> into the <b>Stool Bead Tube</b> .  |
|-------------------|----------|--|
|                   | 3.<br>4. | Vortex to mix and incubate at 70 °C for 5 minutes.<br>Homogenise the sample with maximum speed using cell disruptor<br>(e.g., Digital Disruptor Genie, 2850 rpm) for 20 minutes.           |
| S                 | 5.       | Centrifuge at 8,000 x g for 2 minutes.   |
| Lysis             | 6.       | Transfer <b>500 <math>\mu</math>L supernatant</b> into a new 1.5 mL microcentrifuge tube (not provided).   |
|                   | 7.       | Add <b>150 µL STL2 Buffer</b> and vortex to mix for 5 seconds.   |
|                   | 8.       | Incubate on ice (0 – 4 °C) for 5 minutes.  |
|                   | 9.       | Centrifuge at 16,000 x g for 3 minutes.  |
|                   | 10.      | Place an Inhibitor Removal Column (purple ring) into a 2 mL<br>Microcentrifuge Tube.   |
| Inhibitor Removal | 11.      | Transfer <b>500</b> $\mu$ L clear supernatant into the Inhibitor Removal Column and centrifuge at 16,000 x g for 1 minute.   |
|                   | 12.      | KEEP THE FLOW-THROUGH and discard the column.<br><b>Note:</b> If pellet is observed in the flow-through, transfer the clear supernatant to a new 2 mL microcentrifuge tube (not provided). |
|                   | 13.      | [Optional] Perform RNase treatment if RNA-free DNA is required. Add <b>4</b> $\mu$ L RNase A (not provided) and incubate for 5 minutes at room temperature.                                |



|         | 14. | Add <b>800 µL STL3 Buffer</b> to the flow-through and immediately shake  |
|---------|-----|--|
|         |     | vigorously for 5 seconds.  |
|         | 15. | Short spin to bring down the lysate.   |
| Binding | 16. | Place a <b>PrimeWay Stool Column</b> (green ring) into a new <b>Collection</b><br><b>Tube</b> .  |
| Bin     | 17. | Transfer <b>up to 700</b> $\mu$ L <b>lysate</b> into the <b>PrimeWay Stool Column</b> and centrifuge at 16,000 x <i>g</i> for 1 minute. Discard the flow-through and place the column back into the <b>Collection Tube</b> . |
|         | 18. | Repeat Step 17 until all the lysate has been transferred to the <b>PrimeWay Stool Column</b> .   |
| ۵۵      | 19. | Add <b>400</b> $\mu$ L STL3 Buffer to the PrimeWay Stool Column. Centrifuge at 16,000 x g for 30 seconds. Discard the flow-through and place the column back into the Collection Tube.                                       |
| Washing | 20. | Add <b>600 <math>\mu</math>L Wash Buffer ST</b> to the <b>PrimeWay Stool Column</b> . Centrifuge at 16,000 x <i>g</i> for 30 seconds. Discard the flow-through and place the column back into the <b>Collection Tube</b> .   |
|         | 21. | Repeat Step 20.  |
| 50      | 22. | Centrifuge the column at 16,000 x g for 3 minute to dry the column membrane.   |
| Drying  |     |  |



|         | 23. Transfer the <b>PrimeWay Stool Column</b> to a new 1.5 mL microcentrifuge tube (not provided).   |  |  |
|---------|--|--|--|
| Elution | Add 30 – 100 μL preheated Elution Buffer (60 °C) to the centre of<br>the PrimeWay Stool Column membrane. Incubate at room<br>temperature for at least 2 minutes. Centrifuge at 16,000 x g for<br>2 minutes to elute the DNA. |  |  |
|         | 25. Store the eluted DNA at $2 - 8 \degree$ C or $-20 \degree$ C for long-term storage.  |  |  |



| Problems       | Possible Reason       | Recommended Action                                |
|----------------|-----------------------|---|
| Low DNA Yield  | Too many sample       | Too little space available for beads beating. Use |
| or no recovery |                       | the recommended/ lesser amount of stool.          |
|                | Did not preheat the   | Preheat the Elution Buffer at 60 °C before        |
|                | Elution Buffer        | Elution Step. Ensure the Elution Buffer is        |
|                |                       | completely absorbed by the membrane.              |
|                | Inappropriate buffer  | Ensure the correct amount of absolute ethanol     |
|                | preparation           | is added to the Wash Buffer ST before use.        |
|                |                       | Ensure no precipitation formed in STL1 Buffer.    |
|                |                       | Dissolve the precipitate by incubating in a 37 °C |
|                |                       | water bath, followed by gentle shaking.           |
| Degraded       | Inappropriate storage | Upon stool collection, stool samples should be    |
| DNA            | condition             | kept at 2 – 8 °C if the extraction process is     |
|                |                       | intended to be carried out on the same day. If    |
|                |                       | DNA extraction is not performed on the same       |
|                |                       | day, store the stool in –20 °C until further      |
|                |                       | processing.                                       |
|                | Harsh mechanical      | Reduce disruption (homogenisation) speed and/     |
|                | disruption            | or time.  |
| No PCR         | Presence of PCR       | Dilute the DNA to reduce concentration of PCR     |
| amplification  | inhibitors            | inhibitors.                                       |
| DNA floats     | Presence of ethanol   | In the Drying step, extend the centrifugation     |
| out of agarose | residual              | time to 5 minutes to ensure the PrimeWay Stool    |
| gel well       |                       | Column is completely dry.                         |
| during loading |                       |   |

#### **Troubleshooting Guidelines**

Please contact us at <u>https://base-asia.com/contact/</u> for more information.

#### **Product Ordering Information**

| Product Number   | Product Description   | Remarks                                 |
|------------------|---|---|
| BUF-3024-1X100ml | 1X Tris-EDTA (TE) Buffer, pH 8.0,<br>Biotechnology Grade, 100ml | For dry, hard or high fibre stool only. |
| K.RGT-9110-1ml   | RNase A Solution, 100mg/mL,<br>1mL                              | Optional if RNA-free DNA is required.   |



