

# Product Information

## exTEN II PCR Master Mix (2X), 20 reactions (SAMPLE)

<b>C/No.</b>	BIO-5188-20
<b>Lot No.</b>	-
<b>Expiry Date</b>	-
<b>Concentration</b>	0.06U/μl exTEN II DNA Polymerase, 400μM dNTP mix, 3mM MgCl <sub>2</sub>
<b>Packaging</b>	1 x 500μl
<b>Supplied with</b>	-
<b>Storage</b>	-20°C Avoid frequent thawing and freezing



### Unit Definition

One unit of the enzyme catalyzes the incorporation of 10nmol of deoxyribonucleotides into an acid-insoluble material in 30mins at 74°C by using activated salmon sperm DNA as a template / primer.

### Applications

- DNA sequencing
- DNA labeling
- PCR for cloning
- PCR amplification of DNA fragments up to 12kb
- Routine PCR

### exTEN II PCR Master Mix (2X) Composition

Contains 0.06U/μl exTEN II DNA Polymerase, 400μM dNTP Mix, 3mM MgCl<sub>2</sub>, reaction buffer and a PCR enhancer.

### Description

exTEN II PCR Master Mix (2X) is a unique premixed blend of Taq II DNA Polymerase, a proofreading enzyme, dNTPs, MgCl<sub>2</sub> and reaction buffer for amplification of DNA templates by PCR. With the addition of a 3'→5' exonuclease (proofreading) ability, the amplification efficiency is enhanced through a lower error rate of misincorporated nucleotides compared to just Taq II DNA Polymerase alone. exTEN II PCR Master Mix (2X) produces higher yields and amplifies longer fragments up to 12kb. Most of the amplified DNA fragments have 3'A overhang, while a small percentage are blunt-ended. This premix formulation saves times and reduces contamination by reducing the number of pipetting steps for PCR setup. exTEN II PCR Master Mix (2X) consists of a density reagent and 2 tracking dyes which migrates at the same rate as a 4000bp and 50bp DNA fragment in a 1% agarose gel.

### Quality Control

#### Nuclease Assay

No detectable contaminating endonuclease or exonuclease activity.

#### Functional Assay

exTEN II PCR Master Mix (2X) was tested for amplification of 12000bp of human genomic DNA.

Certified by:

**Ng Xue Qi**

Laboratory Officer

## Protocol

The following protocol serves as a starting point and general guideline for any Polymerase Chain Reaction (PCR). Reaction conditions such as incubation time and annealing temperature may vary and need to be optimized. PCR reactions should be prepared in a DNA-free environment, dedicated pipette and aerosol resistant tips are recommended. Always keep the control and template DNA isolated from other components.

### Recommendations of Template DNA in a 50 µl reaction volume

DNA Type	Amount of DNA
Human genomic DNA	0.1 to 1 µg
Plasmid DNA	0.5 to 5 ng
Phage DNA	0.1 to 10 ng
<i>E.coli</i> genomic DNA	10 to 100 ng

1. Thaw exTEN II PCR Master Mix (2X) at room temperature. Keep the tube on ice after thawing. Vortex briefly and spin down contents quickly.
2. Prepare the following reaction mix in a sterile, nuclease-free PCR tube on ice.

For a 50µl reaction volume:		
Components	Volume (µl)	Final Conc.
exTEN II PCR Master Mix (2X)	25	1X
Forward Primer (10µM)	0.5 – 5	0.1 – 1µM
Reverse Primer (10µM)	0.5 – 5	0.1 – 1µM
DNA template	1 – 5	See above
Nuclease-Free water	Top up to 50	N.A

3. Cap tubes and spin down contents briefly.
4. Place reactions in thermal cycler and incubate at 95°C to completely denature template DNA. Perform about 25 - 35 cycles of PCR amplification by repeating steps 2 - 4 in the PCR reaction. Low amounts of starting template may require 40 cycles.

### Recommended PCR Cycling Condition

No.	Step	Temperature	Time
1	Initial Denaturation	95°C	2 - 4 min
2	Denaturation	95°C	30 seconds
3	Annealing	42 - 65°C	30 seconds
4	Extension	68°C* / 72°C	**0.5-1 min/kb
5	Final Extension	68°C* / 72°C	5 – 10 min
6	Soak	4°C	∞

\*For >5kb amplification, it is recommended to use 68°C as the extension temperature.

\*\*For >3kb amplification, it is recommended to use 1kb/min as extension rate

## General guidelines for PCR amplifications

### Initial denaturation

To ensure efficient utilization of the template during first amplification cycle, it is essential that the template is denatured completely. If GC content of the template is ≤ 50%, an initial 1-5 mins denaturation at 95°C would be sufficient.

### Denaturation

DNA denaturation time of 0.5min/cycle at 95°C is usually sufficient. For GC-rich templates, denaturation could be prolonged to 3-4mins.

### Annealing

Annealing temperature should be 5°C lower than melting temperature ( $T_m - 5^\circ\text{C}$ ) of primers. 0.5min/cycle is usually sufficient. If non-specific PCR products are observed, the temperature should be optimized stepwise in 1-2°C increments.

### Extension

Optimal temperature for extension of exTEN II DNA Polymerase is between 70-75°C. Recommended extension step is 30sec/kb at 72°C for PCR products.

### Number of cycles

If less than 10 copies of template are present, about 40 cycles are required. For higher amount, 25-35 cycles are sufficient.

### Final extension

After the last cycle, incubate PCR mixture at 72°C for an additional 5-15 mins to allow any incomplete amplification to take place.